Flight loss linked to faster molecular evolution in insects

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The loss of flight ability has occurred thousands of times independently during insect evolution. Flight loss may be linked to higher molecular evolutionary rates because of reductions in effective population sizes ($N_e$) and relaxed selective constraints. Reduced dispersal ability increases population subdivision, may decrease geographical range size and increases (sub)population extinction risk, thus leading to an expected reduction in $N_e$. Additionally, flight loss in birds has been linked to higher molecular rates of energy-related genes, probably owing to relaxed selective constraints on energy metabolism. We tested for an association between insect flight loss and molecular rates through comparative analysis in 49 phylogenetically independent transitions spanning multiple taxa, including moths, flies, beetles, mayflies, stick insects, stoneflies, scorpionflies and caddisflies, using available nuclear and mitochondrial protein-coding DNA sequences. We estimated the rate of molecular evolution of flightless (FL) and related flight-capable lineages by ratios of non-synonymous-to-synonymous substitutions ($d_N/d_S$) and overall substitution rates (OSRs). Across multiple instances of flight loss, we show a significant pattern of higher $d_N/d_S$ ratios and OSRs in FL lineages in mitochondrial but not nuclear genes. These patterns may be explained by relaxed selective constraints in FL ectotherms relating to energy metabolism, possibly in combination with reduced $N_e$.

1. Introduction

The evolution of flight in insects is a key innovation thought to have in part enabled their diversification [1–3]. Despite the advantages associated with flight, such as the ability to disperse widely and forage, flight has been independently lost within nearly every pterygote order and an estimated thousands of times overall [4,5]. The lower dispersal ability associated with flight loss tends to increase population subdivision [6,7], with population subdivision expected to reduce the effective population size ($N_e$) of species [8]. Lower dispersal ability may additionally reduce geographical range size (reviewed in [9]) and cause greater fluctuations in population size over time owing to increased susceptibility of (sub)populations to extinction, both of which could limit the $N_e$ of a species [10].

Lineages that differ in $N_e$ over the long term are expected to differ in non-synonymous-to-synonymous substitution ($d_N/d_S$) ratios [11,12] owing to the greater strength of genetic drift in smaller populations. Non-synonymous mutations are more likely to have a fitness effect than synonymous mutations, and the majority of mutations that occur are thought to have a neutral or a deleterious effect [11,13]. In smaller populations, slightly deleterious mutations are more easily fixed, because purifying selection is not as effective at eliminating them, and thus can lead to increased fixation of non-synonymous mutations relative to synonymous mutations. Higher non-synonymous ($d_N$) rates may also produce observable increases in overall substitution rates (OSRs) in those lineages. Therefore, if flightless (FL) lineages did have consistently reduced $N_e$, flightlessness may be a predictor of increased $d_N/d_S$ ratios and OSRs.

A few previous studies of marine organisms, although including a small number of independent comparisons, have used dispersal ability as the predictor of $N_e$ differences and have indeed observed differences in $d_N/d_S$ ratios [14,15]. Molecular patterns have been examined for bird flight loss [16], with the...
expectation of reduced \(N_e\) for FL bird lineages, partially as FL birds are more often found on islands. Additionally, Shen et al. [16] expected and observed higher \(dN/dS\) ratios in FL bird energy-related genes due to a hypothesized relaxation of selective constraints on those genes following the loss of flight. Although the biology of birds and insects differ greatly (e.g. endothermy versus ectothermy), flying metabolic rate in insects is estimated at 50 times higher than resting rate [17,18], and thus we may also observe similar patterns in insects relating to relaxed constraints on energy metabolism. We would expect energy-related influences to mainly affect mitochondrial and nuclear genes involved in the oxidative phosphorylation (OXPHOS) pathway and effective population size influences to have a genome-wide effect.

In this study, we tested for differences in patterns of molecular evolution between FL and flight-capable (F) insect groups by comparing \(dN\) rates, synonymous (\(dS\)) rates, \(dN/dS\) ratios and OSRs of available protein-coding genes. Although there are many ecological or biological factors associated with flight loss (reviewed in [1]), drawing comparisons from a broad range of taxa may help to reduce biases towards certain life-history or ecological traits that could affect the pattern of molecular substitution rates. We have conducted a test of the effect of flightlessness on the rates and patterns of molecular evolution using 49 phylogenetically independent transitions (table 1; [1,5–7,19–36]) that span a wide range of taxa (moths, flies, beetles, mayflies, stick insects, stoneflies, scorpionflies and caddisflies) and different nuclear and mitochondrial protein-coding genes. We approached the analysis in two complementary ways so as to include as many data as possible: whole-tree analysis (as in [14]), which allows analysis of all transitions within a phylogenetic tree and increases the power for detecting a pattern within a tree, and sister-pair analysis (as in [37]), which allows the investigation of individual transitions as independent data points to assess the consistency of any overall trend. Our results show a significant pattern of higher \(dN/dS\) ratios and OSRs across multiple instances of flight loss for mitochondrial genes but not nuclear genes. These results are consistent with our initial hypotheses, although are more favourable towards the idea of relaxed selective constraints on energy metabolism.

2. Material and methods

(a) Data

We collected published molecular phylogenetic studies including both FL and F insect species (see table 1 and the electronic supplementary material, text S1 for description of categorization). We avoided including comparisons where another obvious transition potentially affecting population size or molecular rates occurred simultaneously or subsequent to flight loss (e.g. island living and parasitism) to test ‘flightlessness’ itself while minimizing confounding factors. Phylogenetic relationships were taken from the source studies, with preference for those constructed by model-based methods. Transitions to flightlessness were taken from source studies (see the electronic supplementary material, table S2) assuming flight was never regained [38]. Source phylogenies for whole-tree analysis were trimmed in some cases to eliminate clades that were not suitable for inclusion (e.g. owing to severe confounding factors such as parasitism).

Datasets were constructed as per two methods of analysis; whole tree and sister clade. Overall, 49 independent cases of flight loss were included in the analyses, each represented by at least one protein-coding gene. Eight of the source studies contained two or more independent losses of flight, which include both panmictic and reciprocally monophyletic relationships between F and FL relatives. These phylogenies represented 42 independent transitions in total. These source trees were analysed with both whole-tree and sister-clade methods. For whole-tree analysis, there were 18 trees represented by one gene each. Whole-tree gene data were not combined per study and gene type (nuclear versus mitochondrial) since different transitions were represented by different genes, except in one case where data for two mitochondrial genes were available for identical taxa. In this latter case, the data were combined using \(dN\) and \(dS\) substitution counts and numbers of sites as described further below. Not all of the cases of flight loss within a tree could be included in the sister-clade analysis since sister FL and F pairs had to be matched without overlap of the branches that connect the comparisons on the phylogeny. Thirty-three sister-clade comparisons were obtained from the whole-tree sets, with other studies contributing genetic data towards some of these transitions, and an additional seven transitions were obtained from other studies including single transitions.

Protein-coding gene sequences were downloaded from GenBank, obtained from the authors directly or downloaded from the authors’ webpages (GenBank accession nos. are given in the electronic supplementary material, table S1). Nucleotide sequences were aligned in MEGA v. 5.0 [39] using the Clustal function (default settings) and verified by visual inspection and amino acid translation. Whole-tree alignments were included if they possessed at least two amino acid substitutions in the ingroup and if variability occurred between FL: F pairs. Sister-clade \(dN/dS\) datasets were included if the ingroup (FL→F lineages) contained amino acid differences, or if the calculated non-synonymous (\(dN\)) rate was not 0 for both FL and F lineage types. Sister-clade relative-rate datasets were included if possessing any nucleotide variability between sister lineages. In total, there were 35 \(dN/dS\) sister comparisons and 39 relative-rate sister comparisons that met the criteria for analysis; each transition was represented by at least one gene result (one to seven genes).

(b) \(dN/dS\) ratio and overall substitution rate analysis

\(dN/dS\) ratio analysis and OSR analysis for whole trees, as well as \(dN/dS\) ratio analysis for sister clades, were conducted using maximum-likelihood methods in the program PAML [40]. Component \(codeml\) was used to examine \(dN\), \(dS\) and \(dN/dS\) ratios, and component \(baseml\) was used to examine OSRs; the same input data were used for the two types of whole-tree analyses. All input and output files for all analyses were deposited in the Dryad Digital Repository (doi:10.5061/dryad.3ps4r). For OSR analysis, the sequence models were estimated in MEGA v. 5.0 using constrained topology (maximum likelihood methods, Bayesian Information Criterion). For whole-tree analysis, only lineage tips were coded to two foreground classes: FL or F, with a background rate for all deeper nodes. We applied this method of coding for whole-tree analysis to avoid biasing the results; \(dN/dS\) rates are estimated as higher for tip lineages [41], and deeper nodes tend to be reconstructed as F due to the unidirectional nature of flight loss [38,42].

FL and F lineages in sister-clade \(dN/dS\) codings were each assigned a class from their point of divergence onward by coding the entire pathway of branches leading to taxa, not just lineage tips. \(dN\), \(dS\) and \(dN/dS\) rates and branch lengths were calculated for each FL and F group of lineages in each dataset. Multiple gene results for the same transition were combined per transition, with nuclear and mitochondrial results separately combined since both the \(dN\) and \(dS\) rates of mitochondrial genes are expected to be higher than those of nuclear genes. The combination was performed by multiplying the \(dN\) rate (site-wise estimate) by the number of non-synonymous (N) sites, and
Table 1. Source data for cases of insect flight loss. (The electronic supplementary material gives the species names and GenBank accession numbers, assigned flight status of species and flight state sources. EF-1α, elongation factor 1α; Wgl, wingless; IDH, isocitrate dehydrogenase; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; RpS5, ribosomal protein S5; CAD, carbamoyl-P synthetase/aspartate transcarbamylase/dihydroorotase; H3, histone 3; NF1, neurofibromin; COI, cytochrome c oxidase subunit I; COII, cytochrome c oxidase subunit II; cytB, cytochrome b; FFL, female flightless; bsFL, both-sexes-flightless.)

<table>
<thead>
<tr>
<th>order, taxonomic level and group, ‘common name’</th>
<th>type of flight loss, potential biological associations</th>
<th>no. transitions in tree (no. sister comparisons)</th>
<th>sequences</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lepidoptera, subfamily Ennominae, ‘winter moths’ [19]</td>
<td>FFL, forest dwelling, winter flight, polyphyly</td>
<td>7 (7)</td>
<td>EF-1α, Wgl, IDH, GAPDH, RpS5, COI</td>
</tr>
<tr>
<td>Lepidoptera, tribe Operophterini, ‘winter moths’ [20]</td>
<td>FFL, forest dwelling, winter flight, polyphyly</td>
<td>2 (2)</td>
<td>EF-1α, Wgl</td>
</tr>
<tr>
<td>Lepidoptera, genus Thyrocoa [21]</td>
<td>bsFL, windswept areas</td>
<td>2 (2)</td>
<td>EF-1α, Wgl, COI</td>
</tr>
<tr>
<td>Coleoptera, family Lampyridae, ‘fireflies’ [22]</td>
<td>FFL, female neoteny, loss of male spermatophore</td>
<td>4 (4)</td>
<td>COI</td>
</tr>
<tr>
<td>Coleoptera, family Geotrupidae, ‘earth-boring dung beetles’ [23]</td>
<td>bsFL, arid or semi-arid conditions</td>
<td>6 (5)</td>
<td>NF1, COI, COII</td>
</tr>
<tr>
<td>Mecoptera, order Mecoptera, ‘scorpionflies/hangingflies’ [26],[27]c</td>
<td>bsFL and FFL</td>
<td>4 (4)</td>
<td>EF-1α, COI, COII</td>
</tr>
</tbody>
</table>

aOne of the three transitions in this study overlaps with [19] and the transition was included in whole-tree analyses for this study [20] for one gene that was not available in [19].
bGene did not have enough amino acid variation, but was included in substitution rates tests.
cStudy or gene has overlapping transition with a whole-tree study, adds gene data to sister-clade analysis.

<table>
<thead>
<tr>
<th>order, taxonomic level and group, ‘common name’</th>
<th>type of flight loss, potential biological associations</th>
<th>no. transitions in tree (no. sister comparisons)</th>
<th>sequences</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lepidoptera, tribe Psychidae, sp. [33]</td>
<td>FFL, change from plants to sand insect</td>
<td>1</td>
<td>COI</td>
</tr>
<tr>
<td>Thysanoptera, clade within genus</td>
<td>case material</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lepidoptera, genus Orgyia [30,31]</td>
<td>FFL, spring feeding [32]</td>
<td>1</td>
<td>DDC, EF-1α, CAD, IDH, MDH, RpS5b, COI</td>
</tr>
<tr>
<td>Lepidoptera, family Psychidae, sp. [33]</td>
<td>FFL</td>
<td>1</td>
<td>COI</td>
</tr>
<tr>
<td>Coleoptera, genus Silpha (within ‘carrion beetles’) [17]</td>
<td>bsFL, shift in feeding habit from necrophagy to predatory [34]</td>
<td>1</td>
<td>COI</td>
</tr>
<tr>
<td>Ephemeroptera, genus Cheironomus [35]</td>
<td>bsFL, lack of fish predation [36]</td>
<td>1</td>
<td>H3b</td>
</tr>
<tr>
<td>Plecoptera, Zelandoperla fenestrate [6]</td>
<td>bsFL</td>
<td>1</td>
<td>H3b, COII</td>
</tr>
</tbody>
</table>

Higher variance in rates is expected for sister-clade pairs with low information content [44] and with higher divergences [45]. Since we use a binomial test for analysis of patterns and not a weighted test we consider low information content in the data. All statistical tests performed are 2-tailed.

(c) Sensitivity and additional testing

Higher variance in rates is expected for sister-clade pairs with low information content [44] and with higher divergences [45]. Since we use a binomial test for analysis of patterns and not a weighted test we consider low information content in the data. All statistical tests performed are 2-tailed.
are combined per study; out of the six studies containing mitochondrial genes, all six dN/dS results are higher for the FL than F categories (p = 0.0313). The dN/dS ratios of FL lineages in these six comparisons were approximately double those of the F lineages (median FL/F dN/dS ratio was 2.44). Although nuclear tree results (figure 1a, lighter bars) were not combined due to differing transitions in the trees per study, there is no consistent pattern across the six studies that contain nuclear gene results (two positive (FL > F) results, two negative (F > FL) results and two mixed (both positive and negative gene results for the same study)). Mitochondrial gene dN rates were generally, though not significantly, positive across the studies (five of six studies were positive), and nuclear dN results were not consistent across the studies (two positive, two mixed, one negative and one neutral). dS rates were not consistent across studies for mitochondrial genes, though majority negative (five of six studies negative), nor were they consistent for nuclear genes (three positive, two negative and one mixed). All whole-tree results are given in the electronic supplementary material, table S3.

(ii) Sister-clade results

Thirty-nine independent transitions were analysed. Mitochondrial sister-clade dN/dS results (figure 2a) showed a significant pattern; out of 30 independent comparisons, 23 have higher dN/dS ratios in the FL lineages (p = 0.0052). The dN results are also significantly higher in mitochondrial genes with 24 of 30 comparisons being positive (p = 0.0014). dS mitochondrial rates did not show a significant pattern (17 of 30 positive, p = 0.5847). The dN/dS ratios of FL lineages in these 30 comparisons were approximately double those of the F lineages (median FL/F dN/dS ratio was 1.99). The nuclear sister-clade dN/dS results (figure 2b) did not show a consistent pattern (12 of 26 positive, p = 0.8450) and neither did nuclear dN (12 of 26 positive, p = 0.8450) or dS rates (16 of 26 positive, p = 0.3269). COI was the most common gene available for analysis, and patterns for this gene were consistent with those of the overall mitochondrial results; dN/dS and dN were consistently positive (22 of 27 positive, p = 0.0015 and 21 of 27 positive, p = 0.0039, respectively), whereas dS rates were not (14 of 27 positive, p = 1.000). All dN/dS sister-clade gene results are given in the electronic supplementary material, table S4, and the final combined results are organized in the electronic supplementary material, table S5.

(b) Overall substitution rates

(i) Whole-tree results

Mitochondrial whole-tree OSRs (figure 1b, darker bars) were generally positive but not significantly so across the six studies (results in five of six studies were positive, p = 0.2188). Nuclear whole-tree OSR patterns (figure 1b, lighter bars) were also not consistently positive (four of six studies were positive, one negative and one majority negative).

(ii) Sister-clade relative rates test results

Thirty-nine independent transitions were analysed by relative rates tests. The 30 mitochondrial results showed a significant pattern in which FL lineages had higher relative rates (21 of 30 positive, p = 0.0428), whereas the 32 nuclear results did
(c) Sensitivity testing

No significant negative relationship was observed between the square roots of the sum of the branch lengths and the absolute differences in $d_N/d_S$ rates for the sister pairs. Therefore we did not remove any datasets from $d_N/d_S$ sister-comparison binomial pattern testing.

We do not expect that the node-density effect has produced the observed $d_N/d_S$ sister-clade results. Out of the gene datasets in $d_N/d_S$ sister-clade analysis (82 sets) (see the electronic supplementary material, table S4), almost half (40) were balanced comparisons in which the FL and F groups have an equal number of tips. The remainder of datasets were divided between the FL category having a higher number of individuals per clade (20) and the F category having a higher number of individuals per clade (22).

FFL and both-sexes-flightless (bsFL) transitions were equally represented in the number of mitochondrial and nuclear sister-clade $d_N/d_S$ comparisons. These flight loss types did not differ in mitochondrial $d_N/d_S$ patterns (for FFL 11 of 15 transitions were positive versus bsFL 12 of 15 positive ($p = 1.000$)), $d_N$ results (FFL 12 of 15 versus bsFL 12 of 15 ($p = 1.000$)) or $d_S$ results (FFL 7 of 15 versus bsFL 10 of 15 ($p = 0.4621$)). Nuclear sister-clade patterns also did not differ significantly by flight loss type for $d_N/d_S$ results (FFL 8 of 13 positive versus bsFL 4 of 13 positive ($p = 0.2377$)), $d_N$ results (FFL 7 of 13 versus bsFL 5 of
4. Discussion

FL insect groups displayed higher mitochondrial dN/dS ratios significantly more often than their F relatives. This result was consistent across both whole-tree studies and across sister comparisons, with FL dN/dS rates approximately double those in F lineages. Additionally, there was a significant pattern of mitochondrial OSRs being higher in FL lineages across the sister comparisons. The nuclear patterns for both dN/dS ratios and OSRs were not consistent across whole-tree studies or across the sister-clade comparisons.

Effective population size (N_e) differences are expected to act genome-wide; therefore, the difference in mitochondrial and nuclear patterns suggests that N_e differences may not be the sole or main cause of the patterns. N_e may still play a role in the mitochondrial patterns observed. The N_e of mitochondrial genes is inherently fourfold smaller than autosomal mitochondrial genes and be fixed due to drift. Reduced N_e, for example owing to island living, may affect mitochondrial and nuclear genes differently and only cause accelerated rates in mitochondrial genes [46]. Overall however, it does not appear that there are strong N_e differences consistent across the majority of cases of flight loss tested. The various reasons for expecting N_e differences described in the introduction may in fact be counteracted by the increase in fecundity observed for FL groups [47].

The strong pattern across mitochondrial genes is likely at least partially due to a relaxation of selective constraints associated with flight loss, as was observed for bird flight loss [16]. Flight is an energetically costly activity; insect flying metabolic rate is estimated at 50 times higher than that at rest [17,18]. Therefore, the loss of flight may result in decreased selection pressure to maintain efficient energy production. For example, the winged morph of a grasshopper species consumes significantly more energy than the wingless morph [48], and winged individuals of pea aphids show increased transcription levels of genes related to energy production relative to unwinged morphs [49]. Although these studies examined intraspecific differences in flight ability, energy consumption or expression differences might be expected between FL and F species as well. These various reasons for expecting N_e differences described in the introduction may in fact be counteracted by the increase in fecundity observed for FL groups [47].

Complete agreement across all datasets was not an original expectation of this study owing to expected differences in positive or purifying selection across taxa and genes. While there was variability among datasets, our overall consistent patterns are quite remarkable in the light of the possible confounding influences upon molecular rates. In using flight loss as a predictor of molecular rates, if co-occurring factors that were excluded here (parasitism and island living) were instead included, these might enhance the pattern of higher dN/dS ratios in FL groups. More work is needed to delve deeper into the molecular patterns to elucidate additional influences, for example, of positive or purifying selection. Future studies using larger amounts of genetic data for individual transitions could further test for genome-wide patterns and perform genome-wide scans for positive selection in order to identify the types of genes involved in and affected by flight loss.

In conclusion, we have shown that FL insect lineages have higher ratios of non-synonymous-to-synonymous substitutions and higher OSRs than related F lineages for mitochondrial genes but not nuclear genes. While differences in effective population size between FL and F lineages may exist, our results are more favourable to the hypothesis of relaxed
selective constraints relating to energy metabolism in FL groups. This study has brought to light an interesting wide-spread pattern in insect molecular evolution.

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